

## Lead Tetra-acetate Oxidation of *NN*-Dimethylglycine and its Relevance to the Biosynthesis of the Pyrrolidine Ring of Nicotine

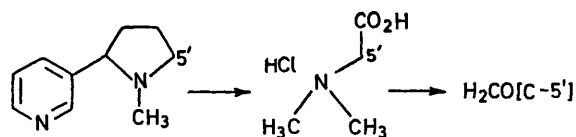
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**Summary** Validity of the degradation of the pyrrolidine ring of nicotine *via NN*-dimethylglycine hydrochloride has been confirmed since oxidation of the latter by lead tetra-acetate yields formaldehyde derived  $98.7 \pm 1.1\%$  from the methylene group.

LEETE, in a recent article in this journal,<sup>1</sup> has criticized our procedure<sup>2</sup> for the degradation of *NN*-dimethylglycine hydrochloride to dimethylamine, formaldehyde, and carbon dioxide. He reports that our oxidation procedure using lead tetra-acetate, in his hands, led to formaldehyde derived from the *N*-methyl groups as well as from the methylene. As support, he cites other observations<sup>3-5</sup> that formaldehyde can be produced by the oxidation of *N*-methyl groups. However, these examples involve ozone<sup>3,4</sup> and periodate<sup>5</sup> as oxidants, not lead tetra-acetate on the amine hydrochloride according to our procedure. He concludes that this contamination from the *N*-methyl group would lead to high values for our reported activity at C-5' (isolated as formaldehyde) in nicotine biosynthesis

with <sup>14</sup>CO<sub>2</sub> and would account for the difference between the unsymmetrical labelling pattern found by us<sup>2</sup> and the symmetrical pattern found by Zielke *et al.*<sup>6</sup>

Apparently Leete overlooked the control experiments we presented<sup>2</sup> which established the validity of our oxidation procedure. One degradation of [2-<sup>14</sup>C]-*NN*-dimethylglycine



hydrochloride we reported as giving formaldehyde with more than 99% of the initial specific activity (now No. 1, Table). The other control degradation was reported with β-<sup>14</sup>C]-β-dimethylaminopropiophenone and ultimately gave formaldehyde with 99.1% of the initial specific activity (now No. 2, Table).

TABLE  
[<sup>14</sup>C]-*NN*-Dimethylglycine,  
HCl<sup>a</sup>

Expt. No.	Labelled carbon	Formaldehyde <sup>b</sup>	
		Spec. act. <sup>c</sup>	% <sup>d</sup>
1	C-2	1.27 × 10 <sup>5</sup>	99.2
2	C-2	57,670	99.1
3	C-2	57,670	99.8
4	<i>N</i> -CH <sub>3</sub>	5.54 × 10 <sup>6</sup>	2.4
5	<i>N</i> -CH <sub>3</sub>	5.54 × 10 <sup>6</sup>	1.4

<sup>a</sup> Prepared by reductive alkylation of glycine with formaldehyde as described for the corresponding  $\beta$ -alanine derivative in ref. 2 and isolated as the hydrochloride by sublimation.

<sup>b</sup> Assayed as its dimedone derivative.

<sup>c</sup> Specific activity in d.p.m./mmol.

<sup>d</sup> Per cent of the original dimethylglycine activity found in the formaldehyde.

We have now performed three additional control degradations, the last two with material prepared from glycine and [<sup>14</sup>C]formaldehyde. All experiments were conducted with

*NN*-dimethylglycine hydrochloride, free of glycine and sarcosine by automatic amino-acid analysis and pure by mass spectrometry, n.m.r., and elemental analyses. The oxidation conditions were as specified<sup>2</sup> and were kept constant throughout the reaction.

As can be seen from the Table, in all the degradations the methylene carbon maintained its integrity to the extent of 98.7 ± 1.1%. The departure from 100% is practically within experimental error (1% of each count) and could have no effect on the reported<sup>2</sup> activities of nicotine at C-5'. Furthermore, this degradation procedure cannot account for the difference between the unsymmetrical<sup>2</sup> and the symmetrical labelling pattern,<sup>5</sup> since in both cases our degradation procedure was used. We have also found symmetrical labelling patterns in some <sup>14</sup>CO<sub>2</sub> biosynthesis experiments, but the factors responsible have not yet been established.

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